PestFacts WA

Issue: 8
Date: June 2025

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Diamondback moths in canola

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- Manypeaks
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- Gibson
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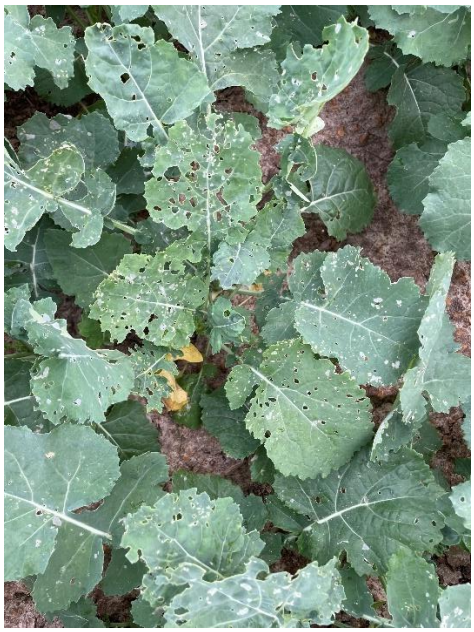
An adult diamondback moth. Photo courtesy of DPIRD.



Diamondback moth caterpillar feeding damage on canola. Photo courtesy of Kieran Zilm (Nutrien).

Kieran Zilm (Nutrien) reports that diamondback moths (DBM) are at threshold levels in a canola paddock at South Stirling. The crop is just at stem elongation. More than 50 DBM caterpillars were found per 10 sweeps and the leaves had shot hole damage.

Small numbers of DBM caterpillars and moths have recently been observed in canola crops at South Stirling, Gibson, and Manypeaks along with the typical 'shot-hole' feeding damage.

The department has deployed pheromone traps to monitor adult moth activity across canola paddocks from Geraldton to Esperance. For more information on this surveillance effort refer to the 2025 PestFacts WA Issue 7 article Timely pest alerts for growers. DBM surveillance is also being conducted at Muresk and Katanning as part of the Grains Research and Development Corporation (GRDC) co-funded Canola Allies Project, in which the Department of Primary Industries and Regional Development (DPIRD) is a partner organisation. To date, the highest number of adult moths have been recorded at Howick (83 moths), where 5 larvae were also found in the crop. Low numbers of DBM (5 or fewer moths) have been detected at Maya, Allanooka, Muresk, Mount Barker and Gibson. No larvae were found in the adjacent canola crop.

Pheromone trap catches indicate the presence of DBM in the landscape. However, DBM larvae are rarely an economic concern early in the season. Populations typically build through late winter and into spring as daytime temperatures increase. For example, at 12°C the DBM life cycle takes over 100 days, while at 28°C it can be completed in just 14 days. As a result, most DBM damage in canola crops tends to occur during spring.

Recent rainfall in some areas is not expected to have reduced numbers of DBM caterpillars, but it may have slowed their lifecycle. Recent GRDC-funded research, conducted by DPIRD, found that DBM moths can colonise canola crops very early, with larval build-up often not seen until temperatures rise at the end of winter (e.g. August). However, above warmer-than-average daytime temperatures are forecast for the WA

grainbelt this winter, which may increase DBM populations in crops. For more climate information, refer to DPIRD's latest Seasonal Climate Outlook.

Research has also found that regular monitoring and recording of DBM numbers, along with changes in larval growth stages (i.e. the percentage of various sized caterpillars) over time, is the best way to anticipate any potential rapid buildup. Tracking changes in numbers of DBM larvae also provides an opportunity to commence spray operations (if required) at a time when they are most likely to be effective.

Identifying diamondback moth caterpillars and their damage

A diamondback moth caterpillar. Photo courtesy of DPIRD.

DBM caterpillars are pale green, cigar-shaped and up to 12 mm in length. They wriggle actively when disturbed and can drop down on a fine thread.

Damage from these caterpillars appears as chewed leaves, buds and flowers, with leaf chewing ranging from irregular holes in leaves to extensive leaf damage.

If you need help with identifying moths or caterpillars, you can take some photos and submit an identification request through the PestFacts WA Reporter app.

Management and considering beneficials

Growers are urged to monitor their crops, correctly identify caterpillar species, and, if crops aren't bouncing back from feeding damage, to consider applying an insecticide. Growers are reminded that insecticides that are effective on DBM can only be applied twice within a season.

Growers and consultants are also advised to monitor for DBM larvae, especially from August onwards, by doing at least four lots of ten sweeps with an insect net at various locations in each crop.

As mentioned previously, DBM caterpillars drop from plants when disturbed, so bashing a few plants, especially those with holes in the leaves, over an ice cream container is a good initial indication of their presence, if you don't have a sweep net handy.

DBM are difficult to control because they breed prolifically, are resistant to many insecticides including synthetic pyrethroids and organophosphates and in advanced canola canopies insecticide sprays have limited coverage.

Thresholds for control are:

- pre-flowering (stressed crop) - 30 or more grubs per 10 sweeps
- pre-flowering (no stress) - 50 or more grubs per 10 sweeps.

Growers are reminded that DBM have shown high levels of resistance to many insecticide groups including synthetic pyrethroids (e.g. alphacypermethrin, esfenvalerate, gamma cyhalothrin, lambda cyhalothrin), carbamates (e.g. methomyl) and organophosphates (e.g. chlorpyrifos). For a list of registered chemicals for DBM in canola, refer to DPIRD's 2025 winter spring insecticide guide.

Growers should consider insecticide options that are soft on predator insects if spraying.

For a list of insecticides with their toxicity to beneficial insects, refer to Cesar Australia's Beneficials Chemical Toxicity Table.

Further information

For more DBM information refer to:

- DPIRD's Diamondback moth and its management in canola and crop weeds factsheet.
- GRDC's Diamondback moth best practice management guide - southern
- GRDC's Managing diamondback moth video.

For more information contact Research Scientist Svetlana Micic in Albany on +61 (0)8 9892 8591.

Article authors: Cindy Webster (DPIRD Narrogin), Dustin Severtson (DPIRD Northam) and Svetlana Micic (DPIRD Albany).

Native budworm moth flights have started

- Ogilvie
- Nabawa
- Maya
- Wongan Hills
- Northam
- York
- Cascade
- Gibson

Native budworm moths captured in a pheromone bucket trap at Nabawa. Photo courtesy of DPIRD.

High numbers of native budworm moths have been trapped north of Geraldton at Ogilvie (136 moths) and Nabawa (98) and east of Carnamah at Maya (53) this week. Smaller native budworm moth numbers were captured in the Esperance shire at Cascade (14 moths) and Gibson (9).

In mid-June, moths were being trapped in the central agricultural region at Wongan Hills (32), the Northam Shire (12,20 and 22) and York (22)

Additional native budworm moth counts can be viewed at Cesar Australia's MothTrapVisWA page, which is regularly updated with the latest trap counts.

Pulse and canola growers in these areas are encouraged to check their crops in the coming weeks as there may be caterpillar offspring from these flights.

This trapping surveillance is part of the "Seasonal status of pests and disease delivered to growers" project, a Grains Research and Development Corporation (GRDC) and Department of Primary Industries and Regional Development (DPIRD) collaboration. For more information refer to the 2025 PestFacts WA Issue 7 article Timely pest alerts for canola growers.

Biology

Native budworm moths lay white spherical eggs (0.5 mm) singly, mostly near the top of the plant. The eggs darken as they mature and tiny caterpillars hatch after about seven days.

The small caterpillars will cause minor leaf damage that is not obvious, such as tiny holes chewed into leaves. Larve can initially remain unnoticed if crops aren't being checked with a sweep-net. The young caterpillars feed on the leaves for about two weeks before they become large enough (5 mm long) to be visible in the crop. However, as the caterpillars get larger, they can cause severe damage.

Management

Growers can inspect plants for caterpillar feeding damage and assess caterpillar numbers using a sweep net, or by bashing plants into a container. This is a good approach if the crop is too short to sweep. Serradella, lucerne, clover and annual medic seed crops should also be regularly checked for native budworm caterpillars.

Pesticide options for the control of native budworm can be found in DPIRD's 2025 Winter Spring Insecticide Guide.

Do you want to host a native budworm trap this season?

From late July to early August volunteer farmers, agronomists, and some DPIRD staff will begin weekly pheromone trapping for native budworm moths. This monitoring program helps to monitor the potential risk of native budworm caterpillars to pulse and canola crops.

For more information refer to the 2025 PestFacts WA Issue 7 article Native budworm moth trapping program will begin in July. Would you like to host a trap?

If you are interested in hosting a trap, please contact Bec Severtson in Northam, or Andrew Phillips in Geraldton.

More information

For more information about the native budworm and its impact on crops refer to the department's Native budworm page.

For further information contact Research Scientists Dusty Severtson in Northam on +61 8 9690 2160, or Andrew Phillips in Geraldton on +61 8 9956 8567.

Article authors: Alan Lord (DPIRD Perth) and Cindy Webster (DPIRD Narrogin).

Article input: Christiaan Valentine, Bec Severtson and Dusty Severtson (all DPIRD Northam).

Lucerne flea activity update

- Binnu
- Yuna
- Nabawa
- Geraldton
- Mooriary
- Dudawa
- Bowgada
- Northam
- Merredin
- Kalgan (Albany)

Northern grain growers, along with Back Paddock and Agworld app users, have reported widespread lucerne flea activity in establishing vetch pastures, wheat, oats, canola and lupin crops ranging from Binnu to Merredin.

Lucerne flea on a lupin plant. Photo courtesy of Alana Hartley (Elders).

Alana Hartley (Elders) has found both lucerne flea and redlegged earth mites damaging a barley crop near Northam.

A lucerne flea and windowing chewing damage on clover seedling. Photo courtesy of James Bee (Elders).

James Bee (Elders) reported extensive lucerne flea damage (to a clover seedling pasture at Kalgan earlier this month. James advised that the crop will be sprayed and resown.

Windowing chewing damage by lucerne flea on canola cotyledon. Photo courtesy of DPIRD.

Lucerne flea hatching, triggered by recent rains, can be patchy across paddocks, and the pest can be more problematic on loam/ clay soils. They attack a range of crops and pastures, causing characteristic ‘windowing’ of leaves. Feeding damage begins at ground level, with the underside of leaves being damaged.

For more information on how to identify lucerne flea damage see DPIRD's 2024 PestFacts WA Issue 7 article Lucerne flea update.

Management of lucerne flea

Lucerne fleas are often patchily distributed in crops, and spot spraying may be sufficient. Lucerne fleas are not effectively controlled by synthetic pyrethroids (SPs).

If crops are not out-growing damage caused by lucerne flea, control relies on application of organophosphates (OPs) such as dimethoate or omethoate. Growers are reminded that chlorpyrifos products can no longer be used on canola, lupin and cereal crops after September 2025.

When both lucerne flea and redlegged earth mites (RLEM) are present, consider control strategies that target both pests. It is recommended to use a product registered for both and apply it at the highest label rate specified for either pest to ensure effective control. If RLEM are resistant to OPs, note that diafenthiuron, which is effective on resistant RLEM, provides suppression of lucerne flea populations in canola.

Crops should be monitored again after spraying as chemical applications will not control lucerne flea eggs that have already been laid. For registered insecticide recommendations for lucerne flea and RLEM refer to DPIRD's 2025 autumn winter insecticide guide.

Further information

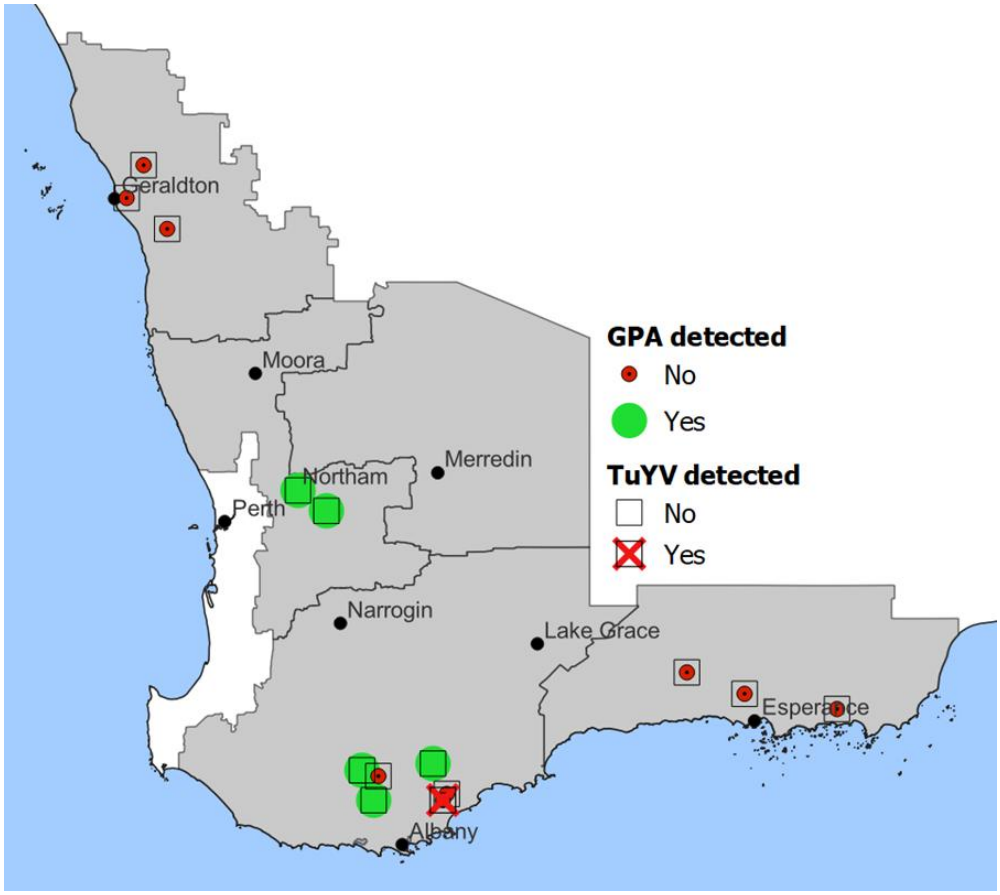
You can request, or confirm, identification of lucerne flea through the PestFacts WA Reporter app or by emailing the PestFacts WA team at pestfactswa@dpird.wa.gov.au.

For more information contact Research Scientists Svetlana Micic in Albany on +61 8 9892 8591 and Andrew Phillips in Geraldton on +61 8 9956 8567.

Article author: Bec Severtson (DPIRD Northam).

Green peach aphid beginning to infest canola crops in Albany and Kwinana West port zones

- Northam
- York
- Cunderdin
- Amelup
- Cranbrook
- Kendenup
- Tenterden
- Albany



Green peach aphid (GPA) and turnip yellows virus (TuYV) detection on yellow sticky traps and canola crops across WA, as of 25 June 2025. Map courtesy of DPIRD.

Plant virology staff from the Department of Primary Industries and Regional Development (DPIRD) found green peach aphid (GPA) infesting less than 10% of canola crops at Amelup, Cranbrook and Kendenup between 11 to 25 June 2025 (see map above). Turnip yellows virus (TuYV) was not detected in either the aphids or the crops. GPA had previously been caught on traps at Amelup in early May. GPA have also been found by Research Scientist Amber Balfour-Cunningham (DPIRD/UWA) in volunteer canola near Tenterden and in wild radish just north of Albany.

In the Kwinana West port zone, GPA was first found infesting less than 10% of plants in a canola crop near York in late May. Since then, winged GPA have been caught on traps

located in Northam, and there have been reports of infestations beginning in crops further east around Cunderdin and north of Meckering. TuYV has not been detected in aphid or plant samples yet, but testing is still in progress.

In the Esperance port zone, no GPA have been found on traps or in canola crops, as many crops are approaching flowering. In the Geraldton port zone, no GPA have been found either, but crops are far less mature and remain at potential risk from GPA infestation and TuYV infection.

Winged and non-winged green peach aphids on canola. Photo courtesy of DPIRD.

Potential symptoms of turnip yellows virus in canola include stunted plant growth and purpling or yellowing of the lower leaves, particularly on leaf margins. Photos courtesy of DPIRD.

The risk of TuYV infection is a high when both GPA and TuYV are present, especially during the early phase of crop development (emergence to 7-leaf stage), when crops are most vulnerable to yield loss. Infection that occurs after stem elongation is unlikely to cause yield losses. As many crops in the Albany region are approaching stem elongation, the risk of TuYV is currently low. However, crops sown later and are still in their early

growth stages should be monitored for GPA, as they are still at risk of infection. The optimal temperature range for GPA population growth is 20-25°C, so cool winter conditions may limit the extent of GPA infestation in some areas over the next month.

DPIRD staff will continue monitoring for aphid activity and TuYV using yellow sticky traps and routine inspections of canola paddocks in the Geraldton, Kwinana West, Albany and Esperance regions until August 2025. This surveillance is co-funded by the Grains Research and Development Corporation (GRDC) project DAW2305-003RTX, “Effective virus management in grains crops”.

Management

The only management strategy available after sowing for TuYV is to use foliar insecticides, which must be applied in the early stages of GPA infestation to be most effective. Routine monitoring from emergence to the 7-leaf stage, involving whole plant visual inspection, must be carried out to ensure a well-timed spray.

To do this, at least 10 plants from the crop must pulled out and closely inspected for GPA, as TuYV can be transmitted by just one or two aphids. GPA are likely to be found near paddock boundaries. If GPA are found, growers are encouraged to contact Research Scientist Benjamin Congdon by emailing Benjamin.Congdon@dpird.wa.gov.au to organise free TuYV testing to assist management decisions.

If infective GPA are found, and infestation rate is still relatively low (e.g. less than 30% of plants infested with infective aphids), a foliar spray may be effective in preventing high levels of spread. Once widespread infestation of infective GPA and subsequent TuYV infection occurs, foliar insecticides will not provide any economic benefit in terms of reducing TuYV inflicted yield losses.

It is important to note that monitoring should occur regardless of whether insecticide seed treatments were applied, as these alone are unlikely to prevent TuYV spread.

For more TuYV management information refer to DPIRD’s Turnip yellows virus and its management in canola factsheet.

Effective chemicals currently available in Australia for control of GPA are limited as GPA has evolved resistance to many insecticides. For more information see GRDC’s Green peach aphid – best practice management guide and Aphid and insecticide resistance management in grain crops.

For registered insecticide recommendations, refer to DPIRD’s 2025 winter spring insecticide guide.

Further information

For more information about GPA, and earlier seasonal activity, refer to the 2025 PestFacts WA Issue 6 article Green peach aphid and turnip yellows virus detected, Issue 5 article No green peach aphid detected yet in DPIRD monitoring and Issue 1 article Enhancing aphid and virus control in canola: beyond seed treatments.

For further information contact Research Scientist Benjamin Congdon, in Perth via email at Benjamin.Congdon@dpird.wa.gov.au.

Article author: Benjamin Congdon (DPIRD Perth).

Sclerotinia apothecia found

- Northam

Sclerotinia apothecia (circled in red). Photo courtesy of (DPIRD).

Apothecia have been found in the Department and Primary Industries and Regional Development (DPIRD) sclerotia depot at Northam this week. They have germinated from sclerotia. It is a warning that the cool moist weather in recent weeks has suited the commencement of the sclerotinia disease lifecycle.

Growers in areas with a history of sclerotinia are reminded to consider their sclerotinia risk when their canola, lupin and pulse crops are close to or at flowering. The disease needs to be managed before sclerotinia symptoms are observed.

Symptoms

Sclerotinia infection on a canola leaf (left) and stem (right). Photos courtesy of: DPIRD.

Sclerotinia infection on a lupin pod. Photo courtesy of DPIRD.

Sclerotinia canopy infection is the most common infection pathway with aerial ascospores produced by apothecia infecting petals during crop flowering. Lesions occur in the upper half of the plant's main stem or branches and can be observed on leaves and pods.

Basal sclerotinia stem rot infection displayed as fluffy white fungal growth at the base of a lupin stem. Photo courtesy of DPIRD.

Basal sclerotinia is a different infection pathway where sclerotia germinate myceliogenically and directly infect the stem at ground level in very wet conditions/seasons. White cottony-looking growth girdles the stem, causing the plant parts above the lesion to wilt and die. DPIRD surveys have found that this is more common in lupin and other pulse crops than in canola.

Management

Currently no foliar fungicides are registered or recommended for reducing basal sclerotinia infection in canola or lupin.

DPIRD research has shown that regular rainfall and high humidity (>75%) in the 3 weeks before and after commencement of flowering are most conducive for damaging levels of canopy sclerotinia to occur in crops. While fungicide application reduces disease severity, it does not necessarily give a yield response, so it's important to consider crop risk and value of disease management carefully each season. For canola crops, consider whether a response to upper canopy blackleg is also likely (refer to DPIRD's UCI BlacklegCM - Blackleg upper canopy infection management app).

Growers need to consider the following factors to determine their risk of sclerotinia and which paddocks to prioritise:

- rotation history of the paddock
- history of sclerotinia in the current paddock and those surrounding it
- rainfall events before and after flowering
- crop growth stage
- dense crops with early canopy cover on loamy soil types are at higher risk.

The SclerotiniaCM decision support tool is available for use by canola growers during flowering to help determine the likely economic returns from applying fungicide at a specific time during flowering for the control of sclerotinia stem rot. The user can specify individual paddock data/history as well as recent and expected weather conditions so that the output relates to their own cropping circumstances. The SclerotiniaCM tool can be downloaded from the App Store or Google Play Store and can be used on both phones and tablet devices. For more information refer to DPIRD's SclerotiniaCM decision support tool page.

Several fungicide products are registered for the control of canopy sclerotinia in canola while options in lupin are more limited. Fungicides need to be applied as recommended per product label. Strategic and responsible use of fungicides will reduce the risk of fungicide resistance developing. For more information on registered foliar fungicides for canola and pulses, including lupins, in WA refer to DPIRD's Fungicides page.

Based on the extensive research conducted by DPIRD the following in-season sclerotinia management options are:

- For canola:
  - Apply a single foliar application at 20-50% bloom, provided conditions are favourable for infection before and during flowering. See Table 1 below for recognising bloom stages in canola. Use the SclerotiniaCM tool for guidance.
  - A second fungicide application at 50% bloom is generally only beneficial in seasons with an extended wet period. Fungicides cannot be applied after 50% bloom, but some products have a longer withholding period, so you do need to check the label. Use the SclerotiniaCM tool for guidance.
- For lupin:
  - Aim to apply fungicide from full to late flowering on the main spike to protect main stem pods as they emerge and penetrate the lower canopy

- A range of products are now registered in lupins which can reduce sclerotinia canopy infection e.g., Veritas® Opti and Miravis® Star.

Table 1 Identification of bloom stages in canola (crop average).

Percent bloom	Number of flowers open on the main stem
5%	<5
10%	10
20% (petal drop commences)	11-14
30%	15-20
50% (full bloom, crop is at its brightest)	>20

Growers and consultants are encouraged to report to the PestFacts WA service any apothecia finds or disease observations as the season progresses.

Further information

Further information can be found at:

- DPIRD's Sclerotinia stem rot and its management in canola factsheet
- DPIRD's Sclerotinia stem rot and its management in lupins factsheet
- GRDC's Sclerotinia stem rot in canola factsheet.
- GRDC's Lupin sclerotinia disease risk assessment guide factsheet.

For more information on Sclerotinia in canola contact plant pathologists Andrea Hills, Esperance on +61 (0)8 9083 1144, Ciara Beard, Geraldton on +61 (0)8 9956 8504, Kylie Chambers, Northam on +61 (0)8 9690 2151 or Jean Galloway, Northam +61 (0)8 6690 2172.

For more information on sclerotinia in lupins contact plant pathologists Ciara Beard, Geraldton on +61 (0)8 9956 8504, Geoff Thomas, Perth on +61 (0)428 947 287 or Zia Hoque, Perth on +61 (0)422 018 633.

Article authors: Ciara Beard (DPIRD Geraldton), Jean Galloway (DPIRD Northam) and Andrea Hills (DPIRD Esperance).

Article input: Zia Hoque (DPIRD Northam).

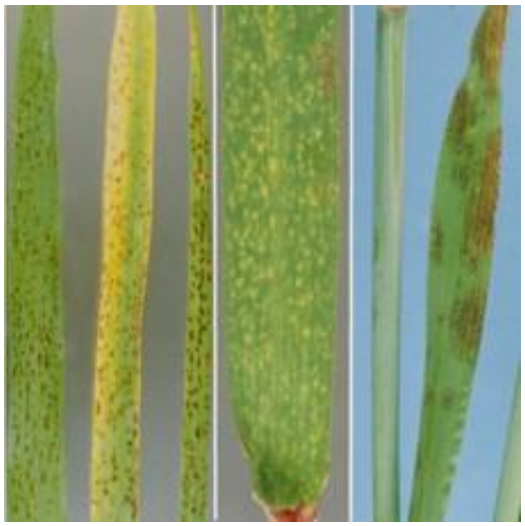
Physiological leaf spotting and chemical damage symptoms can look similar to barley foliar disease

Barley leaves (Neo variety) showing numerous dark spots that are consistent with physiological leaf spotting. Photo courtesy of Aimee Tyson (Central Ag).

Department of Primary Industries and Regional Development (DPIRD) plant pathologists are receiving queries from consultants about barley leaf symptoms, as physiological leaf spotting (PLS) and chemical damage can be confused with foliar diseases such as spot-form net blotch or scald. Sometimes, the similarity in lesions means a diagnosis cannot be determined until lab-based assessments are done, although there are often some distinguishing features.

PLS is a response to an abiotic stress (i.e. not a pathogen) and is not thought to affect yield. PLS spots can develop at any growth stage and are typically distributed in a fairly linear pattern along the leaf blade.

PLS spot symptoms differ between barley varieties. For example, RGT Planet and other varieties with the mlo gene for powdery mildew resistance often display a smudge-like ‘thumb print’, while other varieties tend to show dark spots.



Different physiological leaf spotting symptoms on barley leaves; dark and light speckling (right and centre) and "thumb print" of mlo varieties such as RGT Planet (left). Photos courtesy of: DPIRD.

Physiological leaf spotting on Rosalind barley (left) and Maximus CL barley (right). Photos courtesy of DPIRD.

Early net-form net blotch on barley plants. Lesions have a brown, rectangular pattern. Photos courtesy of DPIRD.

Barley leaf showing mixed symptoms of boron toxicity along the leaf margins and spot-form net blotch. Photo courtesy of DPIRD.

Boron toxicity (on alkaline subsoils) also causes very dark spots, but these are usually, but not always, on the margins of leaves.

Chemical damage in barley plants can be a result of spray drift or crop applications and may present a variety of symptoms. Drift from knockdown herbicides such as paraquat tends to leave a bleached, rounded lesion with a dark margin. As drift is usually a one-off event, the lesions are confined to a single leaf layer and can appear suddenly, whereas net blotch or scald lesions may appear on leaves of various ages and often increase in severity down the canopy. In-crop sprays such as prosulfocarb can cause leaf scorching symptoms.

Neo CL barley plants with lesions from prosulfocarb damage. Photos courtesy of DPIRD.

RGT Planet barley plants with scald lesions of different ages. Newer lesions are a blue-green colour and may lack a margin while older infections become bleached with a dark margin and are unlikely to be spherical like droplet damage. Photo courtesy of DPIRD.

Diagnosis

It is recommended that, where there is any doubt about the cause of cereal leaf spotting, confirmation that it is a fungal disease should be obtained before applying any foliar fungicides. Assistance with disease identification is available through the Department's Diagnostic Laboratory Services - Plant pathology services. There is a fee for this service but can be worthwhile to avoid unnecessary fungicide applications.

Growers can also use the PestFacts WA Reporter app to request a diagnosis of leaf symptoms.

More information

For more information on barley diseases and other causes of leaf spotting see DPIRD's Barley leaf diseases and their management factsheet.

For foliar fungicide information refer to the department's Fungicides page.

For more cereal disease information contact Kithsiri Jayasena, Plant Pathologist, Albany on +61 (0)8 9892 8477, Geoff Thomas, Plant Pathologist, South Perth on +61 (0)8 9368 3262 or Andrea Hills, Plant Pathologist, Esperance on +61 (0)8 9083 1144 or Ciara Beard, Plant Pathologist, Geraldton on +61 (0)8 9956 8504.

Article authors: Cindy Webster (DPIRD Narrogin), Andrea Hills (DPIRD Esperance) and Kithsiri Jayasena (DPIRD Albany).

The PestFacts WA team wants to know what you're finding in the paddock

The PestFacts WA service relies on the WA grains industry to report any invertebrates or diseases found in broadacre crops and pastures – even if you're unsure what they are. In return, we offer diagnoses, management advice and share timely alerts through the PestFacts WA newsletter, map and other communication channels. This helps growers and consultants monitor for emerging issues and respond quickly.

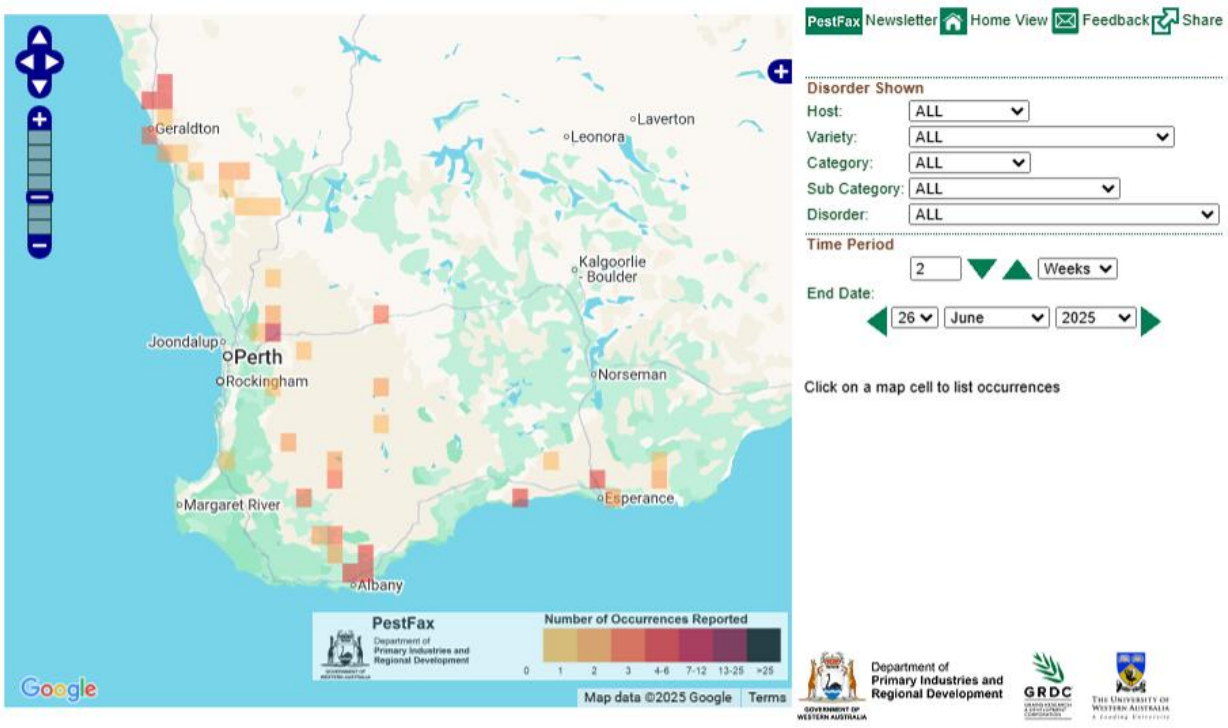
Regular reporting, including noting when pests or diseases are not present, helps build a clearer picture of their distribution and impact. It also supports market access by providing evidence that certain invertebrates and diseases are not present in WA grain production areas.

How do I report or request a diagnosis?

If you find unfamiliar insects in your paddock during the growing season, download and use our PestFacts WA Reporter app to request a diagnosis. Alternatively, you can email the team at pestfactswa@dpird.wa.gov.au

How do I find out what is being found?

Occurrences are visually displayed on DPIRD's PestFacts WA map. This map provides a visual display of all pest and disease occurrences reported by the WA grains industry to the PestFacts WA (formerly PestFax) service since 1996. Viewers can select a host, disorder and time period in the PestFacts WA map.



A screenshot of the PestFacts WA map displaying all disorders reported in all crop and pasture types for the 2 weeks prior to 26 June 2025. Map courtesy of DPIRD.

PestFacts WA newsletters are distributed regularly during the growing season, alerting growers and consultants to insect and plant disease activity, and providing advice on how to manage them. To receive this newsletter directly in your inbox, subscribe here.

Agworld and Back Paddock collaboration

The PestFacts WA team will continue its collaboration with Agworld and Back Paddock this season to collect field data on pests, diseases and weeds. Since 2018 and 2019 respectively, these partnerships have enabled automatic, anonymous reporting from their platforms into the PestFacts WA database via shared Application Programming Interface (API) endpoints.

This collaboration was made possible through a competitive grant process by DPIRD's eConnected Grainbelt project that was initiated in 2017.

If you're interested in participating in a similar collaboration with Agworld, please contact their team at support@agworld.com.

Other collaborative surveillance

The PestFacts WA team collaborates with other DPIRD projects that undertake monitoring and receives regular receives updates from their field observations.

Our work is co-funded with GRDC through the DAW2404-005RTX, Seasonal status of pests and diseases delivered to growers, which carries out routine surveillance for insects and plant diseases. For more detail on the insect surveillance in canola refer to the 2025 PestFacts WA Issue 7 article Timely pest alerts for canola growers during 2025.

All surveillance reports received are published on the PestFacts WA map.

Further information

For more information contact Research Scientist Cindy Webster in Narrogin on +61 8 9881 0201.

Article author: Cindy Webster (DPIRD Narrogin).

Net blotch in barley webinar invitation

Join DPIRD Research Scientists Jason Bradley, Harry Eslick and Jean Galloway for a webinar on Monday 30 June, from 12.30 pm to 1.00 pm (AWST).

The session will cover:

- Managing net blotches in barley
- How to use the new NetBlotchBM decision support tool.

The webinar will be delivered via Microsoft Teams.

To register for this webinar click here.

This webinar is being delivered as part of the GRDC co-investment DAW2112-002RTX, “Disease epidemiology, modelling and delivery of management decision support tools project”.

Article author: Cindy Webster (DPIRD Narrogin).